

available at [www.sciencedirect.com](http://www.sciencedirect.com)journal homepage: [www.ejconline.com](http://www.ejconline.com)

# Granulocyte transfusions in neutropaenic children: A systematic review of the literature

M.D. van de Wetering<sup>a,\*</sup>, N. Weggelaar<sup>a</sup>, M. Offringa<sup>a,b</sup>, H.N. Caron<sup>a</sup>, T.W. Kuijpers<sup>a</sup>

<sup>a</sup>Emma Children's Hospital, Academic Medical Centre (AMC), Amsterdam, The Netherlands

<sup>b</sup>Center for Clinical Epidemiological in Pediatrics, AMC, Amsterdam, The Netherlands

## ARTICLE INFO

### Article history:

Received 26 March 2007

Received in revised form 18 July 2007

Accepted 19 July 2007

Available online 29 August 2007

### Keywords:

Granulocyte transfusions  
Chronic granulomatous disease  
(CGD)  
Paediatric oncology  
Gram-negative bacteraemia  
Fungal infections

## ABSTRACT

**Background:** Granulocyte transfusions (GTX) have been used for decades in paediatric neutropaenic patients, but uncertainty remains regarding their effectiveness. We reviewed all the paediatric data available on GTX, to gain a insight in to the indications for use, favourable effects and side effects in patients and donors.

**Methods:** A comprehensive search was done in MEDLINE, EMBASE, LILACS and CENTRAL (1966 until 2006). All studies including children (1–18 years) who received GTX were included.

**Results:** A total of 66 observational studies were included: Seven using prophylactic and 59 therapeutic GTX. Of the therapeutic studies 55 reported a proven sepsis caused by Gram-negative bacteria (34%) or fungal disease (48%) as the indication for GTX. Concerning effectiveness 70% survival was reported, but no controlled studies were identified. Side effects were mentioned in 27 studies including mild respiratory symptoms, allergic reactions and infection related complications (CMV). Side effects in the donor were mainly flu-like illness.

**Discussion:** In this first review covering 30 years of experience on the use of GTX in children, we found no randomised evidence showing a positive benefit risk ratio. The available case reports and cohort studies alert us as to the potential benefits and harms of the use of GTX in neutropaenic children and provides the basis for a well designed trial in children.

© 2007 Elsevier Ltd. All rights reserved.

## 1. Introduction

Improvement has been made over the past 30 years in cure rates for paediatric oncology patients. This is now estimated around 70%.<sup>1</sup> With the intensification of chemotherapy, the need for adequate supportive care is of utmost importance in maintaining this survival rate. As therapy becomes more intense, infectious complications play a major role. Neutropenia, defined as a neutrophil count of less than 500 cells/ $\mu$ L, is one of the most frequent side effects of childhood cancer treatment. Already in 1966, Bodey et al. showed that the risk

of infections increased rapidly if the neutrophil count dropped below 500 cells/ $\mu$ L and a neutrophil count of less than 100 cells/ $\mu$ L increased the chance of severe infections.<sup>2</sup> Nowadays, of all neutropaenic patients with fever, 12–17% will have a proven bacteraemia or fungaemia.<sup>3</sup> Morbidity due to infections in paediatric oncology patients approaches 30%, and mortality approaches 1%.<sup>4</sup>

Although antibacterials are generally effective, there is a subgroup of patients who do not respond and whose clinical condition deteriorates. For control of infections, granulocytes are crucial. Granulocyte transfusions (GTX) seem a logical and

\* Corresponding author. Tel.: +31 20 566 3050; fax: +31 20 691 2231.

E-mail address: [m.d.vandewetering@amc.uva.nl](mailto:m.d.vandewetering@amc.uva.nl) (M.D. van de Wetering).  
0959-8049/\$ - see front matter © 2007 Elsevier Ltd. All rights reserved.  
doi:10.1016/j.ejca.2007.07.018

potentially effective solution to prevent or overcome longstanding neutropenia and its associated morbidity. GTX have been given to patients as long as 70 years ago.<sup>5</sup> Because of the adverse effects, especially pulmonary reactions and the lack of proven efficacy, GTX almost disappeared from clinical practice for several years. Due to improvement in technology, the interest in GTX has been revived. Three aspects are important, the dose of granulocytes obtained by leukapheresis, the efficiency of the collection procedure, and the donor's neutrophil count.

Granulocytes are now routinely collected by continuous flow centrifugation leukapheresis, and for acceleration of red cell sedimentation, increasing the efficiency of separation a starch and citrated anticoagulant are added.<sup>6</sup> With the availability of recombinant granulocyte-colony-stimulating factor (G-CSF) combined with corticosteroids (oral dexamethasone) higher mean granulocyte yields can be achieved in healthy donors. The granulocyte yield increased 2–10 times compared with the granulocyte yield of controls.<sup>7,8</sup> In addition, after preparation of the donor with G-CSF, the granulocytes showed a better yield and a prolonged half-life of the cells transfused into the recipient,<sup>9</sup> potentially reducing the number of transfusions needed. Side effects were reduced by introducing pre-collection screening of the donor for viral infections (hepatitis, CMV, HIV, EBV). In some critical cases, HLA-cross matching of the donor and the recipient resulted in less sensitisation for other blood products or granulocyte alloimmunisation and reduced the complications of graft-versus-host disease (GVHD).<sup>10,11</sup>

Two meta-analyses were performed in the nineties,<sup>12,13</sup> retrieving studies on GTX published between 1970 and 1994. Both studies concluded that GTX were effective for treatment of severe infection if there was a low survival rate of the controls (<40%), if an adequate number of granulocytes was transfused ( $>10 \times 10^9$ ), and if compatibility of granulocytes was assessed prior to transfusion. However, all the GTX studies performed previously differ in many aspects: the variety of patients included, indications for use, supportive care measures, granulocyte collection procedures and transfusion methods. This makes it very difficult to interpret the data.

Two Cochrane reviews have recently been published.<sup>14,15</sup> In the review of Mohan et al.<sup>14</sup> 4 RCT's were analysed including 44 neonates. Three trials using a small number of patients randomised patients to GTX or no transfusion and one study randomised to GTX or immunoglobulin. Metaanalysis was possible but showed a high percentage of heterogeneity between the trials. The results showed no evidence for use of GTX in septic neonates. All studies transfused a relatively low dose with a mean of  $0.3\text{--}1.0 \times 10^9/\text{kg}$ . In these studies, 4% pulmonary complications were recorded. Stanworth et al.<sup>15</sup> included 8 RCT's; 4 trials included children but the paediatric data were not presented separately. In this review, all but the neonatal RCT's were considered with a total of 310 patient episodes. The relative risk (RR) for mortality extracted from 6 studies was 0.64, but if only studies were included which administered granulocyte counts  $>10 \times 10^9$ , mortality was significantly reduced (RR = 0.37).

To gain an insight into the indications for use, the possible favourable effects of GTX in children and the Side effects for both patient and donor, we reviewed all published paediatric data available on its use.

## 2. Patients and methods

### 2.1. Identification of studies

All relevant literature was searched for, regardless of language or publication status. An internet search was done using the search terms; neutropenia, neutrophil, granulocyte and transfusions. Our search was not limited to children aged 1 year to 18 years, as it was realised that many adult studies included children in their data. If paediatric data within larger studies could be separately analysed they were included in our review. The databases searched were the Cochrane Central Register of Controlled Studies (CENTRAL), published in The Cochrane Library (Issue 4, 2006) MEDLINE (1966 to August 2006), EMBASE (1980 to August 2006), LILACS (1982 to August 2006) and The Web of Science (until August 2006).

### 2.2. Inclusion/exclusion criteria

We included randomised and quasi-randomised controlled studies, phase II trials, cohort, case-control studies, case reports and data from abstracts that included children aged 1 year to 18 years with a malignancy or immunological disorder, neutropenia and evidence of infection (proven bacteraemia, proven fungal sepsis or clinical strong suspicion) or children having an increased chance for infection. Studies using GTX from all sources of granulocytes and using all different methods of collection both for therapeutic or prophylactic purpose were included. We excluded studies giving GTX to neonates, as this was covered in a separate Cochrane review.<sup>14</sup>

### 2.3. Data extraction and outcome measures

Two reviewers (MvdW, TK) independently abstracted the following data: type of study, year of publication, characteristics of the patients, including age and type of malignancy, the number of patients with bacteraemia, the number of patients with fungaemia and the number of patients with a strong clinical suspicion of infection. The number of days of neutropenia (granulocytes  $\leq 500$  cells/ $\mu\text{L}$ ), the mean number of granulocytes transfused, the response of the infectious episode (survival or death) were collected. Side effects in the patient such as TRALI (transfusion related acute lung injury), the insidious onset of pulmonary insufficiency manifested by severe dyspnoea, hypoxia and radiographical evidence of pulmonary oedema with normal cardiac function,<sup>16</sup> platelet or erythrocyte sensitisation, allergic reaction to granulocyte concentrates, and transmission of infections (HIV, hepatitis B or C virus CMV, EBV) were recorded. Data recorded on the donor were stimulation of the donor using G-CSF, corticosteroids, or both, and side effects seen in the donor. The method of collection of granulocytes was documented. Disagreements with regard to the correct data to be extracted were resolved between the two reviewers by discussion.

### 2.4. Data analysis

The data were summarised in a descriptive analysis.



**Table 2 – Reports <1984 use of granulocyte transfusions in oncological disorders**

Study	Patients (#)	Organism	Mean # GTX (granulocytes)	Donor	Side effects	Outcome
Pole et al. <sup>31</sup>	Haematological (30)	Bacterial (Gram-positive, n = 12; Gram-negative, n = 10) fungal (n = 2)	$20 \times 10^9$ + HES, $7 \times 10^9$ – HES	Not stimulated	Pulmonary (3)	25 Survival, 5 death
Schmitz-Valkenberg et al. <sup>32</sup>	Haematological (1)	Bacterial	2 ( $23 \times 10^9$ total)	Not stimulated		1 Survival
Alavi et al. <sup>33</sup>	Haematological (10)	Bacterial (Gram-negative, n = 4)	3–18 ( $33 \times 10^9/\text{m}^2/\text{d}$ )	Steroids	Allergic reactions (3); fever (1)	5 Survival, 1 death
Salfner et al. <sup>32</sup>	Haematological (1)	Pneumonia	2 ( $65 \times 10^9$ )	Steroids	Allergic reaction (1)	1 Death
Rosen et al. <sup>34</sup>	Haematological (1)	Bacterial (Gram-positive, n = 1; Gram-negative, n = 1)	4	Steroids		1 Death
Berkman et al. <sup>35</sup>	Haematological (28)	Bacterial	$(21 \pm 1.3 \times 10^9/\text{m}^2/\text{d})$	Not stimulated		25 Survival, 3 death
Fernandez et al. <sup>36</sup>	Haematological (7)	Bacterial (n = 5) fungal (n = 2)	Range 6–25 ( $1\text{--}5 \times 10^9/\text{m}^2/\text{d}$ )	Steroids		5 Survival, 2 death
Pflieger et al. <sup>37</sup>	Haematological (1)	Bacterial (Gram-negative)	21 ( $63 \times 10^9/\text{m}^2/\text{d}$ )	Not stimulated		1 Death
Estes et al. <sup>38</sup>	Haematological (1)	Fungal	1 ( $60 \times 10^9$ total)	Not stimulated		1 Death
Ritchey et al. <sup>39</sup>	Haematological (5)	Bacterial (Gram-positive, n = 3; Gram-negative, n = 1)	7 ( $4\text{--}20 \times 10^9$ total)	Not stimulated		5 Survival
Dana et al. <sup>40</sup>	Haematological (1)	Fungal	1 ( $13 \times 10^9$ total)		Dyspnoea during GTX	1 Death
Pflieger et al. <sup>41</sup>	BMT (15)	Bacterial (Gram-negative, n = 5) fungal (n = 1)	5 ( $47 \times 10^9/\text{m}^2/\text{d}$ )	Steroids		14 Survival, 1 death
Graubner et al. <sup>42</sup>	Haematological (10)	Bacterial Gram-negative (n = 3)	7, range 6–12... ( $21 \times 10^9$ total)	Steroids		7 Survival, 3 death
Kulkarni et al. <sup>43</sup>	Haematological (1)	Fungal (n = 1)	5			1 survival
Winton et al. <sup>44</sup>	Haematological (20)		( $16 \times 10^9$ total)	Steroids		14 Survival, 6 death
Schmidmeier et al. <sup>45</sup>	Lymphoma (1)		( $10\text{--}15 \times 10^9$ total)	Not stimulated	Allergic reaction (n = 2)	1 Death

**Table 3 – Reports >1984 use of granulocyte transfusions in immune disorders**

Study	Patients (#)	Organism	Mean # GTX (granulocytes)	Donor	Side effects	Outcome
Fanconi et al. <sup>46</sup>	CGD (2)	Bacterial (Gram-positive, n = 1) fungal (n = 1)	5 ( $4 \times 10^9$ total)	Not stimulated		2 Survival
Depalma et al. <sup>47</sup>	CGD (1)		46 ( $16 \times 10^9$ total)	steroids		1 Survival
Drakos et al. <sup>48</sup>	Aplastic anaemia (1)	fungal (n = 1)		Not stimulated		1 Death
Lekstrom-Himes et al. <sup>49</sup>	CGD (1)	Bacterial (Gram-positive)	8		Allergic reaction	1 Survival
Stroncek et al. <sup>50</sup>	CGD (11)		8 ( $20 \times 10^9$ total)	Steroids and G-CSF	Pulmonary (7) & allergic reaction (11)	
Van Planta et al. <sup>51</sup>	CGD (1)	Bacterial (Gram-positive)	8 ( $49 \times 10^9$ total)	G-CSF		1 Survival
Ozsahin et al. <sup>52</sup>	CGD (1)	Bacterial (Gram-negative) & fungal	4 ( $30 \times 10^9$ total)	G-CSF		1 Survival
Van't Hek et al. <sup>53</sup>	CGD (1)	Fungal	5			1 Survival
Girmenia et al. <sup>54</sup>	Aplastic anaemia (1)	Fungal	39 ( $2 \times 10^9$ /kg/d) ( $50 \times 10^9$ /m <sup>2</sup> /d)	G-CSF		1 death
Notarangelo et al. <sup>55</sup>	CGD (1)	Fungal	7	?	Headache, fever	1 Survival
Ikinciogullari et al. <sup>56</sup>	CGD (3)	fungal	8 ( $0.4-3 \times 10^9$ /kg/d)	steroids and G-CSF		2 Survival, 1 death

CGD = chronic granulomatous disease, G-CSF = granulocyte colony stimulating factor.

**Table 4 – Reports >1984 use of granulocyte transfusions in combined immune and oncology disorders**

Study	Patients (#)	Organisms	Mean # GTX (granulocytes)	Donor	Side effects	Outcome
Saarininen et al. <sup>57</sup>	Haematological (9), SAA (1)	Bacterial (Gram-positive, n = 4; Gram-negative, n = 3) fungal (n = 3)	38 ( $59 \times 10^9$ total) ( $0.1-1 \times 10^9$ /kg/d)	G-CSF	None	10 Survival
Parco et al. <sup>58</sup>	Haematological (3) thalassaemia (1)	Bacterial (Gram-negative, n = 2) fungal (n = 1)	8 ( $40 \times 10^9$ total)	G-CSF		3 Survival
De Montalembert <sup>59</sup>	BMT (41), other (4)		4 ( $0.9 \times 10^9$ /kg/d)	Steroids	Pulmonary (1)	26 Survival, ? death
Grigull et al. <sup>60</sup>	Haematological (3) SAA (1)	Fungal (n = 2)	7.5 ( $50 \times 10^9$ total) ( $1.5 \times 10^9$ /kg/d)	G-CSF	Pulmonary (1)	2 Survival, 2 death
Lemau de Talance et al. <sup>61</sup>	Haematological (4) CGD (7)	Fungal (n = 8)	8 ( $30 \times 10^9$ total)	Steroids and G-CSF		10 Survival, 1 death
Sharon et al. <sup>62</sup>	CGD (1) Blackfan-Diamond (1) ALL (1)	Bacterial (Gram-positive; n = 1) fungal (n = 2)	9 ( $5 \times 10^{10}$ /m <sup>2</sup> /d)	Steroids and G-CSF	GvHD	3 Survival

BMT = bone marrow transplant, CGD = chronic granulomatous disease, G-CSF = granulocyte colony stimulating factor, SAA = severe aplastic anaemia.

**Table 5 – Reports >1984 use of granulocyte transfusions in oncology disorders**

Study	Patients (#)	Organisms	Mean # GTX (granulocytes)	Donor	Side effects	Outcome
Yamada et al. <sup>63</sup>	haematological (7)	?	3, range 2–5 (13.6 ± 5 × 10 <sup>9</sup> /m <sup>2</sup> /d)	?	40% Unexpected fever, no serious complications	?
Sciorelli et al. <sup>64</sup>	Haematological (30)	?	3 (19 × 10 <sup>9</sup> total)	Steroids		27 Survival 3 death
Dutcher et al. <sup>65</sup>	Haematological (1)	Fungal (n = 1)	4 (25 × 10 <sup>9</sup> total)	Steroids		1 Survival
Engelhard et al. <sup>66</sup>	Haematological (2)	Bacterial (Gram-negative, n = 2)	3.5 (0.5 × 10 <sup>9</sup> /kg/d)	Not stimulated		2 Survival
Barrios et al. <sup>67</sup>	BMT (1)	Fungal (n = 1)				1 Survival
Angel et al. <sup>68</sup>	Haematological (16)	Bacterial (Gram-negative, n = 16)	2			11 Survival 5 death
Duncan et al. <sup>69</sup>	Haematological (2)	Bacterial (Gram-negative, n = 3)				2 Survival
Morrison et al. <sup>70</sup>	BMT (1)	Fungal (n = 1)	3			1 Survival
Bhatia et al. <sup>71</sup>	BMT (50)	Fungal (n = 50)	14, Range 4–20... (10 × 10 <sup>9</sup> total)	Steroids and G-CSF	Pulmonary	17 Survival 33 death
Rabodonirina <sup>72</sup>	Haematological (6)	Fungal (n = 6)				4 Survival 2 death
Murphy et al. <sup>73</sup>	Haematological (1)	Bacterial (Gram-negative, n = 1)		G-CSF	None	1 Survival
De Silvestro et al. <sup>74</sup>	Haematological (5)		8 (47 × 10 <sup>9</sup> total)			4 Survival 1 death
Bielorai et al. <sup>75</sup>	Haematological (1)	Bacterial (Gram-positive, n = 1)	(48–68 × 10 <sup>9</sup> total)	G-CSF		1 Survival
Price et al. <sup>8</sup>	BMT (4)	Bacterial (Gram-negative, n = 1) fungal (n = 4)	12	Steroids and G-CSF	Chills (37%), temp rise (32%) and itching (11%)	1 Survival 3 death
Lee et al. <sup>76</sup>	Haematological (2)	Bacterial (Gram-negative, n = 2)	2 (66 × 10 <sup>9</sup> total)	Steroids and G-CSF	Pulmonary	1 Survival 1 death
Johnston et al. <sup>77</sup>	Haematological (4)	Bacterial (Gram-negative, n = 3) fungal (n = 1)	5	Steroids and G-CSF		3 Survival 1 death
Illerhaus et al. <sup>78</sup>	Haematological (3)	Bacterial (Gram-negative, n = 1) fungal (n = 1)	5 (26 × 10 <sup>9</sup> total)	G-CSF	Infection (CMV)	2 Survival 1 death
Vij et al. <sup>79</sup>	BMT (2)		2	G-CSF	Infection (CMV)	
Cesaro et al. <sup>80</sup>	Haematological (13)	Bacterial (Gram-positive, n = 5; Gram-negative, n = 8) fungal (n = 4)	4 (52 × 10 <sup>9</sup> total)	G-CSF	Pulmonary	7 Survival 6 death
Lin et al. <sup>81</sup>	Haematological (1)	Bacterial (Gram-negative, n = 1)	10 (32 × 10 <sup>9</sup> total)	G-CSF		1 Survival
Lee et al. <sup>82</sup>	Haematological (32)	Bacterial (Gram-positive, n = 12; Gram-negative, n = 18) fungal (n = 15)	4 (76–92 × 10 <sup>9</sup> total)	Steroids and G-CSF	Pulmonary	10 Survival 1 death
Yoshihara et al. <sup>83</sup>	Haematological (2)	Bacterial (n = 2)	4 (0.7–2.1 × 10 <sup>9</sup> /kg/d)	Steroids and G-CSF		2 Survival
Meyer-Koenig et al. <sup>84</sup>	Haematological (1)	Bacterial (Gram-negative) & fungal	3	G-CSF	CMV	1 death
Ansari et al. <sup>85</sup>	Haematological (1)	Fungal (n = 1)	?	Steroids and G-CSF		1 Survival
Grigull et al. <sup>86</sup>	Haematological (3)	Fungal (n = 3)	9, range 6–15	G-CSF	Fever & chills after 5 GTX	3 Survival
Grigull et al. <sup>87</sup>	Haematological (32)	Bacterial (Gram-positive, n = 6; Gram-negative, n = 3) fungal (n = 6)	3, range 1–19 (2 × 10 <sup>9</sup> /kg/d)	Steroids and G-CSF	Small % fever, resp distress, hypotension and erythema	13 Death (8 because of infection)
Kikuta et al. <sup>88</sup>	Haematological (13)	Bacterial (Gram-positive, n = 3; Gram-negative, n = 8) fungal (n = 2)	2, range 1–4 (0.6 × 10 <sup>9</sup> /kg/d)	G-CSF	Transient hypoxia (n = 2)	9 Survival, 2 death
Sachs et al. <sup>89</sup>	Haematological (26) aplastic anaemia (1)	Bacterial (Gram-positive, n = 2; Gram-negative, n = 3) fungal (n = 7)	2 (0.8 × 10 <sup>9</sup> /kg/d)	Steroids and G-CSF	Minor transfusion reactions (2)	25 Survival

CMV = cytomegalovirus.



Table 6 – Number of patients recorded with infection

	Therapeutic GTX-onco 37 studies	Therapeutic GTX-imm. 13 studies	Therapeutic GTX combined 5 studies	Total therapeutic GRX	Prophylactic GTX-onco 3 studies	Prophylactic GTX-imm 1 study	Prophylactic GTX-combined 0 studies	Total prophylactic GTX
Gram positive sepsis	44 Patients	5 Patients	4 Patients	53 Patients	0 Patients	0 Patients	0 Patients	0 Patients
Gram negative sepsis	97 Patients	1 Patient	5 Patients	103 Patients	4 Patients	1 Patient	0 Patients	5 Patients
Fungal	108 Patients	19 Patients	14 Patients	141 Patients	3 Patients	1 Patient	0 Patients	4 Patients

studies combined these patients (9%). In the 44 oncology studies and the 6 combined studies, a total of 456 patients were included of whom the majority were haematological patients ( $n = 340$ ; 74.5%), and all other patients were allogeneic bone marrow transplant (BMT) patients ( $n = 116$ ; 25.4%). In the 16 studies with CGD patients and in the 6 combined studies 54 patients were included.

Fifty-nine studies reported the exact age of the included patients (mean age of 8.9 years, range 2–17 years), and 7 studies reported age as <18 years.

### 3.3. Infections

In the 59 studies using GTX therapeutically, 55 studies reported a proven sepsis. There were 53 patients (18%) who received granulocytes because of a Gram-positive bacteraemia, 103 patients (34%) received GTX because of a Gram-negative bacteraemia and 141 patients (48%) received this for proven fungal disease. In the 7 studies using prophylactic granulocyte transfusions 4 studies reported a proven sepsis, 5 patients had a previous Gram-negative bacteraemia, and 4 patients had previous fungal disease. The GTX were used as secondary prophylaxis.

The numbers of oncology and immunology patients reported with a proven sepsis is reported in Table 6. Gram negative infections and fungal infections were mainly recorded as a cause for intervening with GTX; however Gram positive infections were also recorded. All these studies reported that the patient's condition did not improve on adequate antibiotic treatment and adequate supportive care management. These Gram positive infections were for example *Staph. aureus* hepatic abscesses leading to a sepsis, or *Streptococci* infections leading to an ARDS. In all described oncology patients with a proven sepsis the mean duration of granulocytopenia was 13 days (range: 2–30 days).

### 3.4. Granulocyte transfusions

In the studies using therapeutic GTX, a mean number of 8.7 granulocyte concentrates were administered (range: 1–46). In the studies using prophylactic GTX the mean number was 4.8 (range: 2–8). Selecting only the paediatric oncology patients receiving therapeutic GTX, a mean of 5.4 transfusions were administered (range: 1–21).

The mean number of granulocytes transfused in all therapeutic GTX studies was  $32 \times 10^9/L$  ( $2-82 \times 10^9/L$ ), which is a mean of  $2 \times 10^9/kg/day$  (range:  $0.5-7.0 \times 10^9/kg/day$ ). The mean number of granulocytes transfused as prophylactic treatment was  $33 \times 10^9/L$  ( $22-46 \times 10^9/L$ ) which is a mean of  $1.3 \times 10^9/kg/day$  (range:  $0.9-1.8 \times 10^9/kg/day$ ).

### 3.5. Donor

Data on the donor were reported in 51 (45 therapeutic studies on GTX and 6 prophylactic use of GTX) out of 66 studies. Out of 45 studies using therapeutic GTX 20 (44.4%) reported the use of related donors, 24 (53.6%) used unrelated donors, and one trial used both. Of these donors 23.4% were stimulated with G-CSF, 19.2% stimulated with steroids, 34% were not

stimulated and 23.4% were stimulated with both corticosteroids and G-CSF.

There was a significant difference in the yield of granulocytes if the donors were not stimulated or if they were stimulated with both G-CSF and steroids (mean:  $18.5 \pm 16 \times 10^9/L$  versus  $58 \pm 26 \times 10^9/L$ , respectively;  $p = 0.003$ ). The difference between G-CSF stimulated only and steroid-stimulated only was not significant (mean:  $42 \pm 13 \times 10^9/L$  versus  $30 \pm 17 \times 10^9/L$ , respectively;  $p = 0.132$ ).

In the studies using prophylactic GTX, data from the donor were known in 6 out of 7 studies. Three studies used related donors, and three studies used unrelated donors. Of these 60% were stimulated with G-CSF and 40% were stimulated with steroids without any significant difference in yield.

### 3.6. Graft handling

To increase the granulocyte yield various methods have been used. The use of 6% hydroxyethyl starch (HES) infused continuously during the granulocyte collection was reported in 31 studies out of 66 (67.3%). The procedures used to collect granulocytes have been done with the IBM 2997 cell separator (10%), the Cell Separator 3000 (42%), the COBE cell separator (26%), and a continuous flow cell separator (22%). This was reported in 41 out of 66 included studies. To decrease the risk of Side effects all leucocyte concentrates were irradiated before administration to the patient; that this was actually done was reported in 19 of the included studies.

### 3.7. Clinical outcome

In patients receiving therapeutic GTX, the outcome was recorded in 57 included studies, which includes 11 prospective studies and 46 retrospective studies. In the 31 case reports, 81 patients survived and 53 patients died. In the 21 cohort studies altogether 168 patients survived and 50 died. Of the reported patients 70% showed survival and 30% of patients died.

There was no significant difference in the granulocyte number infused between the patients who survived ( $21 \pm 15 \times 10^9/L$ ) and the patients who died ( $20 \pm 10 \times 10^9/L$ ). However, if one calculated the number of deaths in the group of which the donors were stimulated with G-CSF and steroids compared to no stimulation, 50% less deaths were seen in the stimulated group. There were 43 deaths in the non-stimulated group and 20 deaths reported in the stimulated group.

### 3.8. Side effects

Pulmonary complications in the recipient were reported in 7 of the included studies. This ranged from mild symptoms to more severe respiratory symptoms, TRALI was not reported. This was mostly seen before 1974 when graft handling was essentially different. GVHD was seen in 3 case studies. In these 3 studies from the 197's the transfused granulocytes were not irradiated. In 9 studies allergic reactions were recorded; all these resolved without complications. In 3 studies transmission of infection was reported mainly as CMV infection. Side effects in the donor were mentioned only in a few

studies. Flu-like symptoms were mentioned in 3 studies in donors who were stimulated with G-CSF.

## 4. Discussion

In this review all paediatric data on the use of GTX were recorded. In the time period between 1976 and 2006 this was reported in 510 paediatric patients. All these patients were treated to the best available knowledge on GTX at that time. Yet, no RCT was performed exclusively in children >1 year of age, and it is therefore difficult to estimate the efficacy of GTX in neutropaenic paediatric patients with severe infections. Reported indications for GTX use in this group of patients show that more than 80% of patients received GTX for a proven Gram-negative or fungal infection. The mortality and morbidity for these infections is much higher than for Gram-positive organisms, and, in addition, an increase is seen in recent years in the occurrence of Gram-negative infections.<sup>17–19</sup>

The mean duration of neutropaenia in these described patients was 12.3 days which corresponds with the high risk criteria for febrile neutropenia where >10 days is defined as prolonged neutropaenia.<sup>20</sup> From all data reviewed on paediatric patients GTX might be beneficial in severe neutropaenia of long duration who experience a Gram-negative or fungal sepsis. In these reported paediatric patients 70% of patients survived and 30% of patients died. This does not however reflect the efficacy as positive results could be reported more than negative results. In the recently published Cochrane review on 8 RCTs mainly in adult patients<sup>15</sup> the RR for mortality was only significantly reduced (RR 0.37, 95%CI 0.17–0.82) when studies were considered in which adequate numbers of granulocytes were transfused ( $>10 \times 10^9$ ). None of these RCT's looked at exactly the same outcome, so the authors concluded that these results should be interpreted with caution.

In all the described studies of our review, it was shown that donors stimulated with a combination of corticosteroids and G-CSF had the highest granulocyte yield and in the group of patients receiving granulocytes of prestimulated donors fewer deaths were recorded. This suggests that patients benefit more from donor-prestimulated than unstimulated neutrophils even when receiving equal numbers of cells. This can be best explained by an inherent advantage of G-CSF and/or dexamethasone to the cellular activities of donor neutrophils, such as survival, motility or killing capacity, some of which have been well documented.<sup>21,22</sup>

Concerning Side effects in the patient, the severe pulmonary complications that were seen in the past were not reported in all the case reports and cohort studies described after introduction of novel leukopheresis methods. There were mild pulmonary signs and symptoms that all resolved without late effects. Other complications were GVHD and allergic reactions. After irradiating all granulocyte products the risk of GVHD seems irrelevant. Side effects in the donor were only minimally reported, mainly consisting of flu-like symptoms in the G-CSF stimulated donors. No long term follow-up has been reported of these donors. Safety concerns on administration of G-CSF therapy to healthy donors has been reported. Bennett et al.<sup>23</sup> describe 5 cases (out of 738 donors)



who developed a haematologic malignancy years after G-CSF therapy, a causal relationship cannot be demonstrated but it does stress the need for long term follow-up.

There are no RCT's for the use of GTX in paediatrics. Our review covers 30 years of observational experience and reflects the 'best available evidence'. Weaknesses of our analysis are the retrospective, uncontrolled, non-randomised, non-blinded intervention, the non-blinded outcome assessments, and lack of adequate follow-up. Selection, performance, detection and attrition bias could all play a role and a simple conclusion on the efficacy of the use of GTX in neutropaenic children can not be drawn.

Several authors<sup>24,25</sup> have emphasised the potential importance of observational evidence. It alerts us as to the potential harms or benefits of a treatment. Given the results of this review, there remains doubt on the efficacy of the use of GTX in reducing mortality without increasing morbidity in paediatric neutropaenic patients.

We have learned from this review in children that there should be a clear clinical indication for the use of GTX and that granulocytes should be collected under optimal, donor-prestimulated circumstances.

In conclusion, paediatric neutropaenic patients with a fungal infection or bacterial infection form a target group of patients who could benefit from GTX. We are as yet uncertain on the exact timing and the exact dose of GTX, and are definitely in need of standardised indications and protocols. We recommend that GTX should not be given outside the realm of a well-designed clinical trial.

## Conflict of interest statement

None declared.

## REFERENCES

- Coleman MP, Gatta G, Verdecchia A, et al. EUROCARE-3 summary: cancer survival in Europe at the end of the 20th century. *Ann Oncol* 2003;**14**(Suppl 5):v128–49.
- Bodey GP, Buckley M, Sathe YS, Freireich EJ. Quantitative relationships between circulating leukocytes and infection in patients with acute leukemia. *Ann Intern Med* 1966;**64**:328–40.
- Lucas KG, Brown AE, Armstrong D, Chapman D, Heller G. The identification of febrile, neutropaenic children with neoplastic disease at low risk for bacteremia and complications of sepsis. *Cancer* 1996;**77**:791–8.
- Verhoef J, Verhage EA, Visser MR. A decade of experience with selective decontamination of the digestive tract as prophylaxis for infections in patients in the intensive care unit: what have we learned? *Clin Infect Dis* 1993;**17**:1047–54.
- Strumia MM. The Effect of leukocytic cream injections in the treatment of the neutropenias. *Am J Med Sci* 1934;**187**:527–44.
- Vamvakas EC, Pineda AA. Determinants of the efficacy of prophylactic granulocyte transfusions: a meta-analysis. *J Clin Apher* 1997;**12**:74–81.
- Lee JH, Leitman SF, Klein HG. A controlled comparison of the efficacy of hetastarch and pentastarch in granulocyte collections by centrifugal leukapheresis. *Blood* 1995;**86**:4662–6.
- Price TH, Bowden RA, Boeckh M, et al. Phase I/II trial of neutrophil transfusions from donors stimulated with G-CSF and dexamethasone for treatment of patients with infections in hematopoietic stem cell transplantation. *Blood* 2000;**95**:3302–9.
- Leavey PJ, Thurman G, Ambruso DR. Functional characteristics of neutrophils collected and stored after administration of G-CSF. *Transfusion* 2000;**40**:414–9.
- Dutcher JP, Schiffer CA, Johnston GS, et al. Alloimmunization prevents the migration of transfused indium-111-labeled granulocytes to sites of infection. *Blood* 1983;**62**:354–60.
- McCullough J, Weiblen BJ, Clay ME, Forstrom L. Effect of leukocyte antibodies on the fate in vivo of indium-111-labeled granulocytes. *Blood* 1981;**58**:164–70.
- Vamvakas EC, Pineda AA. Meta-analysis of clinical studies of the efficacy of granulocyte transfusions in the treatment of bacterial sepsis. *J Clin Apheresis* 1996;**11**:1–9.
- Strauss RG. Rebirth of granulocyte transfusions: should it involve pediatric oncology and transplant patients? *J Pediatr Hematol Oncol* 1999;**21**:475–8 [Review] [19 refs].
- Mohan P, Brocklehurst P. Granulocyte transfusions for neonates with confirmed or suspected sepsis and neutropaenia. *Cochrane Database Syst Rev* 2003;CD003956.
- Stanworth SJ, Massey E, Hyde C, et al. Granulocyte transfusions for treating infections in patients with neutropenia or neutrophil dysfunction. *Cochrane Database Syst Rev* 2005;CD005339.
- Popovsky MA, Moore SB. Diagnostic and pathogenetic considerations in transfusion-related acute lung injury. *Transfusion* 1985;**25**:573–7.
- Aksu G, Ruhi MZ, Akan H, et al. Aerobic bacterial and fungal infections in peripheral blood stem cell transplants. *Bone Marrow Transplant* 2001;**27**:201–5.
- Gaytan-Martinez J, Mateos-Garcia E, Sanchez-Cortes E, et al. Microbiological findings in febrile neutropenia. *Arch Med Res* 2000;**31**:388–92.
- Haupt R, Romanengo M, Fears T, Viscoli C, Castagnola E. Incidence of septicaemias and invasive mycoses in children undergoing treatment for solid tumours: a 12-year experience at a single Italian institution. *Eur J Cancer* 2001;**37**:2413–9.
- Hughes WT, Armstrong D, Bodey GP, et al. 2002 guidelines for the use of antimicrobial agents in neutropaenic patients with cancer. *Clin Infect Dis* 2002;**34**:730–51.
- Leavey PJ, Sellins KS, Thurman G, et al. In vivo treatment with granulocyte colony-stimulating factor results in divergent effects on neutrophil functions measured in vitro. *Blood* 1998;**92**:4366–74.
- Maiani NA, Mul FP, van Buul JD, Roos D, Kuijpers TW. Granulocyte colony-stimulating factor inhibits the mitochondria-dependent activation of caspase-3 in neutrophils. *Blood* 2002;**99**:672–9.
- Bennett CL, Evens AM, Andritsos LA, et al. Haematological malignancies developing in previously healthy individuals who received haematopoietic growth factors: report from the Research on Adverse Drug Events and Reports (RADAR) project. *Br J Haematol* 2006;**135**:642–50.
- Glasziou P, Vandenbroucke JP, Chalmers I. Assessing the quality of research. *BMJ* 2004;**328**:39–41.
- Vandenbroucke JP. Case reports of suspected adverse drug reactions: case reports were dismissed too quickly. *BMJ* 2006;**332**:488.
- Ippolito RJ, Seashore JH, Touloukian RJ. Excision of pulmonary and renal aspergillomas. Its use in treating chronic granulomatous disease of childhood. *Arch Surg* 1978;**113**:640–2.
- Pedersen FK, Johansen KS, Rosenkvist J, Tygstrup I, Valerius NH. Refractory *Pneumocystis carinii* infection in chronic granulomatous disease: successful treatment with granulocytes. *Pediatrics* 1979;**64**:935–8.

28. Chusid MJ, Shea ML, Sarff LD. Determination of posttransfusion granulocyte kinetics by chemiluminescence in chronic granulomatous disease. *J Lab Clin Med* 1980;**95**:168–74.
29. Cohen MS, Isturiz RE, Malech HL, et al. Fungal infection in chronic granulomatous disease. The importance of the phagocyte in defense against fungi. *Am J Med* 1981;**71**:59–66.
30. Yomtovian R, Abramson J, Quie P, McCullough J. Granulocyte transfusion therapy in chronic granulomatous disease. Report of a patient and review of the literature. *Transfusion* 1981;**21**:739–43.
31. Pole JG, Davie M, Kershaw I, Barter DA, Willoughby ML. Granulocyte transfusion in treatment of infected neutropaenic children. *Arch Dis Child* 1976;**51**:521–7.
32. Schmitz-Valckenberg M, Borberg H. [Therapy with granulocyte transfusions] [German]. *Dtsch Med Wochenschr* 1976;**101**:1458–60.
33. Alavi JB, Root RK, Djerassi I, et al. A randomized clinical trial of granulocyte transfusions for infection in acute leukemia. *N Engl J Med* 1977;**296**:706–11.
34. Rosen RC, Huestis DW, Corrigan Jr JJ. Acute leukemia and granulocyte transfusion: fatal graft-versus-host reaction following transfusion of cells obtained from normal donors. *J Pediatr* 1978;**93**:268–70.
35. Berkman EM, Eisenstaedt RS, Caplan SN. Supportive granulocyte transfusion in the infected severely neutropaenic patient. *Transfusion* 1978;**18**:693–700.
36. Fernandez MN, Barbolla L, Sanjuan I, et al. [Considerations on the use of granulocytes obtained by intermittent flow centrifugation [IFC] in the transfusion to granulocytopenic patients (author's transl)] [Spanish]. *Sangre (Barc)* 1979;**24**:878–95.
37. Pflieger H, Arnold R, Bhaduri S, et al. Beneficial effect of granulocyte transfusions in patients with defects in granulocyte function and severe infections. *Scand J Haematol* 1979;**22**:33–41.
38. Estes SA, Hendricks AA, Merz WG, Prystowsky SD. Primary cutaneous aspergillosis. *J Am Acad Dermatol* 1980;**3**:397–400.
39. Ritchey AK, Andiman W, McIntosh S, Berman B, Luce D. Mononucleosis syndrome following granulocyte transfusion in patients with leukemia. *J Pediatr* 1980;**97**:267–9.
40. Dana BW, Durie BG, White RF, Huestis DW. Concomitant administration of granulocyte transfusions and amphotericin B in neutropaenic patients: absence of significant pulmonary toxicity. *Blood* 1981;**57**:90–4.
41. Pflieger H, Wiesneth M, Arnold R, Niethammer D, Kleihauer E. Granulocyte transfusions in children. *Eur J Pediatr* 1981;**135**:261–6.
42. Graubner M, Kretschmer V, Löffler H, Mueller-Eckhardt C, Pralle H. [Indications and clinical aspects of granulocyte transfusions (author's transl)] [German]. *Dtsch Med Wochenschr* 1981;**106**:1726–32.
43. Kulkarni R, Murray DL, Gupta S, de Mendonca WC, Leader I. Multiple splenic aspergillomas in a patient with acute lymphoblastic leukemia. *Am J Pediatr Hematol Oncol* 1982;**4**:141–5.
44. Winton EF, Moffitt S, Vogler WR, et al. Influence of quantity of granulocytes transfused on response. *Prog Clin Biol Res* 1982;**88**:75–91.
45. Schmidmeier W, Feil W, Gebhart W, et al. Fatal graft-versus-host reaction following granulocyte transfusions. *Blut* 1982;**45**:115–9.
46. Fanconi S, Seger R, Gmur J, et al. Surgery and granulocyte transfusions for life-threatening infections in chronic granulomatous disease. *Helv Paediatr Acta* 1985;**40**:277–84.
47. Depalma L, Leitman SF, Carter CS, Gallin JI. Granulocyte transfusion therapy in a child with chronic granulomatous disease and multiple red cell alloantibodies. *Transfusion* 1989;**29**:421–3.
48. Drakos PE, Nagler A, Or R, et al. Invasive fungal sinusitis in patients undergoing bone marrow transplantation [Review] [31 refs]. *Bone Marrow Transplant* 1993;**12**:203–8.
49. Lekstrom-Himes JA, Holland SM, DeCarlo ES, et al. Treatment with intralesional granulocyte instillations and interferon-gamma for a patient with chronic granulomatous disease and multiple hepatic abscesses. *Clin Infect Dis* 1994;**19**:770–3.
50. Stroncek DF, Leonard K, Eiber G, et al. Alloimmunization after granulocyte transfusions. *Transfusion* 1996;**36**:1009–15.
51. von PM, Ozsahin H, Schroten H, Stauffer UG, Seger RA. Greater omentum flaps and granulocyte transfusions as combined therapy of liver abscess in chronic granulomatous disease. *Eur J Pediatr Surg* 1997;**7**:234–6.
52. Ozsahin H, von PM, Muller I, et al. Successful treatment of invasive aspergillosis in chronic granulomatous disease by bone marrow transplantation, granulocyte colony-stimulating factor-mobilized granulocytes, and liposomal amphotericin-B. *Blood* 1998;**92**:2719–24.
53. van 't Hek LG, Verweij PE, Weemaes CM, et al. Successful treatment with voriconazole of invasive aspergillosis in chronic granulomatous disease. *Am J Respir Crit Care Med* 1998;**157**:1694–6.
54. Girmenia C, Iori AP, Boecklin F, et al. Fusarium infections in patients with severe aplastic anemia: review and implications for management [Review] [17 refs]. *Haematologica* 1999;**84**:114–8.
55. Notarangelo LD, Casa C, Ferrari E, et al. Resolution of pulmonary and vertebral aspergillosis in a child with CGD following combined use of granulocyte transfusions and voriconazole. *ICAAC Abstract Book* 2000:786.
56. Ikinciogullari A, Dogu F, Solaz N, et al. Granulocyte transfusions in children with chronic granulomatous disease and invasive aspergillosis. *Ther Apher Dial* 2005;**9**:137–41.
57. Saarinen UM, Hovi L, Vilinikka L, Juvonen E, Myllyla G. Reemphasis on leukocyte transfusions – induction of myeloid marrow recovery in critically ill neutropaenic children with cancer. *Vox Sang* 1995;**68**:90–9.
58. Parco S, Bruno G, Durighello M, et al. Granulocyte transfusion in leukopenic children by simplified leukapheresis of related donors. *Int J Artif Organs* 1998;**21**:63–4.
59. de Montalembert M et al. Granulocyte transfusion in children. Basic science and clinical practice in blood transfusion, 131b. 1-1-1998. Ref Type: Generic.
60. Grigull L, Schrauder A, Schmitt-Thomssen A, Sykora K, Welte K. Efficacy and safety of G-CSF mobilized granulocyte transfusions in four neutropaenic children with sepsis and invasive fungal infection. *Infection* 2002;**30**:267–71.
61. Lemau DT, Benomar D, Boulat C, Beaumont J-L. Current data on granulocyte donations. *Transfus Clin Biol* 2004;**11**:106–12.
62. Sharon RF, Bierings M, Vrieling H, Versluys B, Boelens JJ. Pre-emptive granulocyte transfusions enable allogeneic hematopoietic stem cell transplantation in pediatric patients with chronic infections. *Bone Marrow Transplant* 2006;**37**:331–3.
63. Yamada T. Critical analysis of granulocyte transfusion in three different methods [filtration leukapheresis (FL), intermittent flow centrifugation (IFC) and continuous flow centrifugation (CFC)]. 2 Clinical efficacies. *Shikoku Acta Med* 1984;**40**:705–15.
64. Sciorelli GA, Ravagnani F, Pellegris G. Granulocyte transfusion therapy. *Int J Artif Organs* 1984;**7**:353–6.
65. Dutcher JP, Kendall J, Norris D, et al. Granulocyte transfusion therapy and amphotericin B: adverse reactions? *Am J Hematol* 1989;**31**:102–8.
66. Engelhard D, Aker M, Or R, Shinar E. Combined therapy with granulocyte transfusion, intravenous opsonins and antibiotics for overwhelming *Pseudomonas aeruginosa*

- septicemia in neutropaenic cancer patients. *Pediatr Infect Dis J* 1989;8:332–4.
67. Barrios NJ, Kirkpatrick DV, Murciano A, et al. Successful treatment of disseminated *Fusarium* infection in an immunocompromised child. [Review] [34 refs]. *Am J Pediatr Hematol Oncol* 1990;12:319–24.
  68. Angel C, Patrick CC, Lobe T, Rao B, Pui CH. Management of anorectal perineal infections caused by *Pseudomonas aeruginosa* in children with malignant diseases. *J Pediatr Surg* 1991;26:487–93.
  69. Duncan BW, Adzick NS, deLorimier AA, et al. Necrotizing fasciitis in two children with acute lymphoblastic leukemia. *J Pediatr Surg* 1992;27:668–71.
  70. Morrison VA, Weisdorf DJ. *Alternaria*: a sinonasal pathogen of immunocompromised hosts. *Clin Infect Dis* 1993;16:265–70.
  71. Bhatia S, McCullough J, Perry EH, et al. Granulocyte transfusions: efficacy in treating fungal infections in neutropaenic patients following bone marrow transplantation. *Transfusion* 1994;34:226–32.
  72. Rabodonirina M, Piens MA, Monier MF, et al. *Fusarium* infections in immunocompromised patients: case reports and literature review. *Eur J Clin Microbiol Infect Dis* 1994;13:152–61.
  73. Murphy JJ, Granger R, Blair GK, et al. Necrotizing fasciitis in childhood. *J Pediatr Surg* 1995;30:1131–4.
  74. De Silvestro G, Marson P, Varotto S, et al. Granulocyte concentrates: new findings. *Trasfusione del Sangue* 1997;42:295–8.
  75. Bielora B, Neumann Y, Avigad I, et al. Successful treatment of vancomycin-resistant *Enterococcus sepsis* in a neutropaenic patient with G-CSF-mobilized granulocyte transfusions. *Med Pediatr Oncol* 2000;34:221–3.
  76. Lee JJ, Chung IJ, Park MR, et al. Clinical efficacy of granulocyte transfusion therapy in patients with neutropenia-related infections. *Leukemia* 2001;15:203–7.
  77. Johnston DL, Waldhausen JHT, Park JR. Deep soft tissue infections in the neutropaenic pediatric oncology patient. *J Pediatr Hematol Oncol* 2001;23:443–7.
  78. Illerhaus G, Wirth K, Dwenger A, et al. Treatment and prophylaxis of severe infections in neutropaenic patients by granulocyte transfusions. *Ann Hematol* 2002;81:273–81.
  79. Vij R, DiPersio JF, Venkatraman P, et al. Donor CMV serostatus has no impact on CMV viremia or disease when prophylactic granulocyte transfusions are given following allogeneic peripheral blood stem cell transplantation [see comment]. *Blood* 2003;101:2067–9.
  80. Cesaro S, Chinello P, De Silvestro G, et al. Granulocyte transfusions from G-CSF-stimulated donors for the treatment of severe infections in neutropaenic pediatric patients with onco-hematological diseases. *Support Care Cancer* 2003;11:101–6.
  81. Lin YW, Adachi S, Watanabe K, Umeda K, Nakahata T. Serial granulocyte transfusions as a treatment for sepsis due to multidrug-resistant *Pseudomonas aeruginosa* in a neutropaenic patient. *J Clin Microbiol* 2003;41:4892–3.
  82. Lee J-J, Song H-C, Chung I-J, et al. Clinical efficacy and prediction of response to granulocyte transfusion therapy for patients with neutropenia-related infections. *Haematologica* 2004;89:632–3.
  83. Yoshihara T, Ishida H, Morimoto A, et al. Granulocyte transfusions for severe infections prior to allogeneic hematopoietic stem cell transplantation. *Rinsho Ketsueki* 2004;45:557–61.
  84. Meyer-Koenig U, Hufert FT, Duffner U, Neumann-Haefelin D, Henschen M. G-CSF-mobilised granulocyte transfusion to an ALL patient complicated by cytomegalovirus transmission. *Bone Marrow Transplant* 2004;34:1095–6.
  85. Ansari M, Ozsahin H, Gervais A, Wacker P, Taylor S. Cutaneous aspergillosis in a child with an acute lymphoblastic leukemia. *Pediatr Blood Cancer* 2005;45:1005–6.
  86. Grigull L, Beilken A, Schmid H, et al. Secondary prophylaxis of invasive fungal infections with combination antifungal therapy and G-CSF-mobilized granulocyte transfusions in three children with hematological malignancies. *Support Care Cancer* 2006;14:783–6.
  87. Grigull L, Pulver N, Goudeva L, et al. G-CSF mobilised granulocyte transfusions in 32 paediatric patients with neutropaenic sepsis. *Support Care Cancer* 2006;14:910–6.
  88. Kikuta A, Ohto H, Nemoto K, et al. Therapeutic transfusions of granulocytes collected by simple bag method for children with cancer and neutropaenic infections: results of a single-centre pilot study. *Vox Sang* 2006;91:70–6.
  89. Sachs UJ, Reiter A, Walter T, Bein G, Woessmann W. Safety and efficacy of therapeutic early onset granulocyte transfusions in pediatric patients with neutropenia and severe infections. *Transfusion* 2006;46:1909–14.